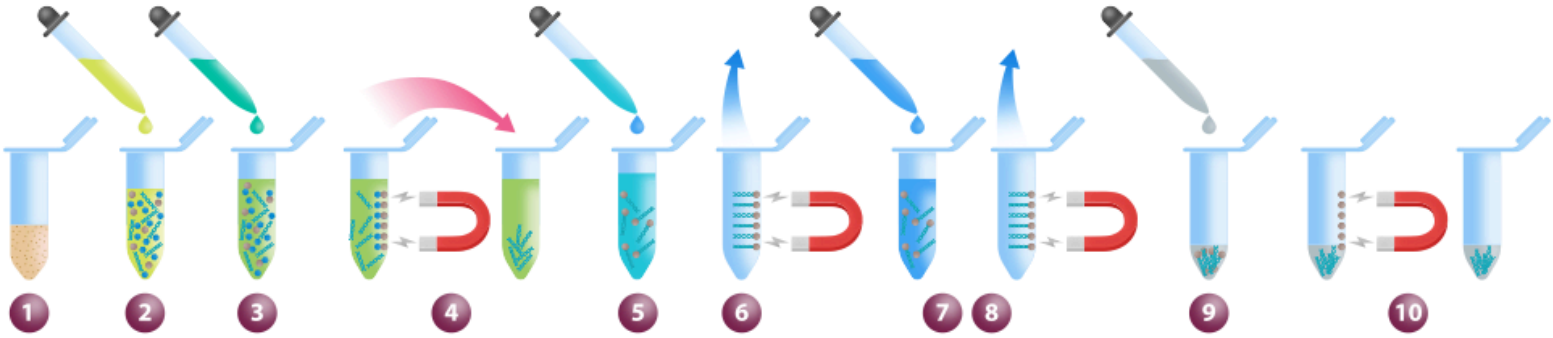
magnetiC

Plasmid DNA Miniprep

Magnetic Bead-Based Plasmid DNA Extraction and Purification



1. Pellet 1 mL of bacterial cell sample. Discard supernatant. Add 200 µL 1X PBS to the pellet and mix well by pipetting up-down until solution is homogenous
2. Add 200 µL of **Plasmid Lysis Buffer** and mix well by pipetting up-down (5-10x) until solution is homogenous. Incubate for 5 minutes at room temperature
3. Add 100 µL of **Neutralization Buffer** and mix well by pipetting up-down (5-10x). Incubate for 5 minutes at room temperature
4. Place microfuge tube on magnetic rack and wait 1 min. Transfer supernatant to new microfuge tube. Discard previous tube.
5. Add 500 µL of **Binding Buffer** to the supernatant in the new microfuge tube and mix well by pipetting up-down (10-15x) until solution is homogenous. Incubate for 1 min at room temperature
6. Place microfuge tube on magnetic rack and wait 1 min. Discard supernatant
7. Add 600 µL of **Wash Buffer** and mix well by pipetting up-down (5-10x) until solution is homogenous. Place microfuge tube on magnetic rack and wait 1 min. Discard supernatant
8. Repeat Step 7 and leave to dry for 1 min
9. Add 25-100 µL of **Elution Buffer** and mix well by pipetting up-down (10-15x) until solution is homogenous. Incubate for 1 min
10. Place microfuge tube on magnetic rack and wait 1 min. Transfer pure plasmid supernatant to a clean microfuge tube

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